

THE STRUCTURE OF ROOSTER-COMB DERMATAN SULFATE. FRAGMENTATION OF THE POLYSACCHARIDE CHAINS BY CHONDROITINASE AC-II DIGESTION, AND BY PERIODATE OXIDATION, FOLLOWED BY ALKALI CLEAVAGE

KINZO NAGASAWA*, AKIRA OGAMO, AND KEIICHI YOSHIDA**

School of Pharmaceutical Sciences, Kitasato University, 9-1, Shirokane 5 chome, Minato-ku, Tokyo 108 (Japan)

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ABSTRACT

The polysaccharide-chain fragments of rooster-comb dermatan sulfates (RC-20 and RC-30) were obtained by chondroitinase AC-II digestion and by periodate oxidation, followed by alkaline cleavage, and their structures analyzed both quantitatively and qualitatively. RC-20 having a lower D-glucuronic acid content (22.6%) is composed preponderantly of large clusters of *N*-acetyldermosine sulfate ($M_r \sim 17\,600$ – $41\,000$) at the nonreducing terminal, whereas RC-30, having a higher D-glucuronic acid content, (41.4%) is poor in this cluster. Both RC-20 and RC-30 have an *N*-acetyldermosine sulfate cluster (M_r 6500–7300) within the polysaccharide chains. Most *N*-acetylchondrosine sulfate units of RC-20 and RC-30 exist as clusters, the large clusters ($M_r \sim 17\,600$) being preponderant in RC-30; both RC-20 and RC-30 contain a large proportion of *N*-acetylchondrosine sulfate clusters (M_r 3500 and 9000) that corresponds to the uronic acid content. In RC-30, most *N*-acetyldermosine disulfate units (13.4%) are linked to *N*-acetylchondrosine sulfate units or clusters.

INTRODUCTION

In the preceding communication¹, it was shown that the two fractions, RC-20 and RC-30, are representative, both quantitatively and qualitatively, of rooster-comb dermatan sulfate and that these polysaccharides ($M_r \sim 41\,000$) show the highest content of D-glucuronic acid ever observed^{2–4}. These fractions were desulfated and depolymerized under solvolytic conditions⁵ in hot dimethyl sulfoxide containing water to give fractions of di-, tetra-, and hexa-saccharides as the main components. Quantitative and qualitative analyses of these fractions suggested that the polysaccharide chains contain many clusters of *N*-acetylchondrosine or *N*-acetyl-

*To whom correspondence should be addressed.

**Chief Research Staff of Tokyo Research Laboratory, Seikagaku Kogyo Co. Ltd.

dermosine sulfate units, especially the latter ones. Further information on the copolymeric structure of the chains was obtained by digestion with chondroitinase AC-II, and by periodate oxidation followed by alkaline cleavage, and the results are described herein.

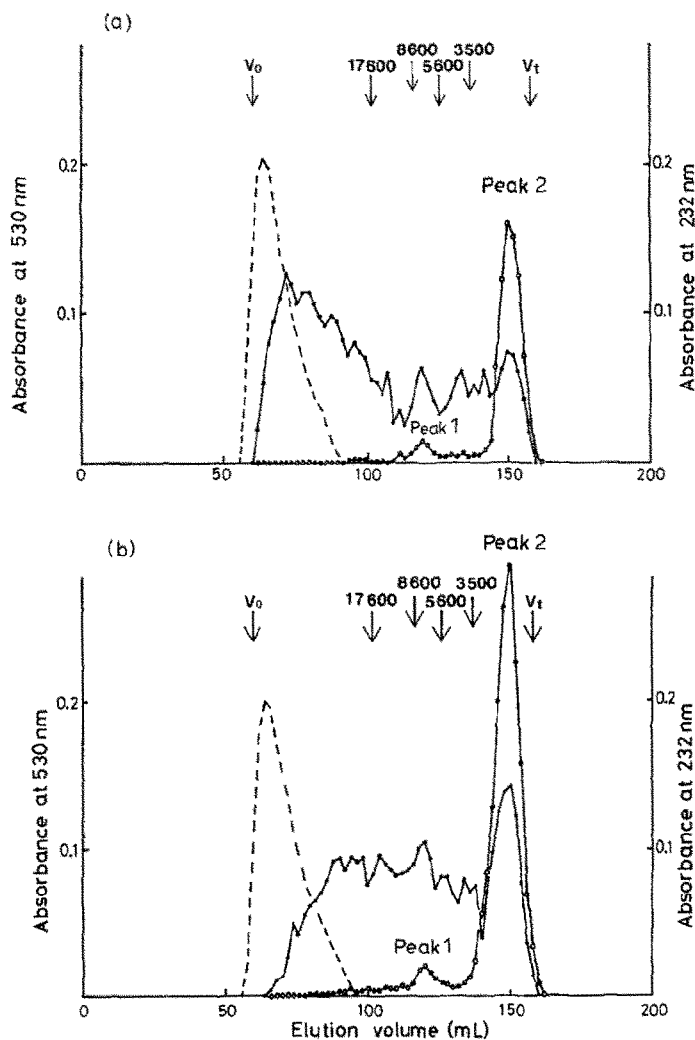


Fig. 1. Separation, on Sephadex G-200, of the products formed by digestion, with chondroitinase AC-II, of rooster-comb dermatan sulfates: (a) Fraction RC-20 and (b) Fraction RC-30. Each column was calibrated with Blue dextran (V_0), depolymerized chondroitin 6-sulfates (M_r 17 600, 8600, 5600, and 3500), and 2M sodium chloride (V_t). (---) Elution diagram of original dermatan sulfate (RC-20 or RC-30), (—●—●—●) $A_{530\text{nm}}$, and (—○—○—○) $A_{232\text{nm}}$.

EXPERIMENTAL

Materials. — The purified fractions (RC-20 and RC-30) of rooster-comb dermatan sulfate were described previously¹. A series of standard chondroitin 6-sulfates (sodium salt) having different M_r values (3500, 5600, 8600, and 17 600) were obtained by limited depolymerization of chondroitin 6-sulfate with bovine testis hyaluronidase and by repeated gel-chromatography of digestion products⁶. Chondroitinase AC-II and chondroitinase ABC were products of Seikagaku Kogyo Co. Ltd., Tokyo.

Analytical methods. — Reducing power was determined by the method of Park-Johnson⁷. Other analytical procedures were carried out as previously described¹.

Digestion of rooster-comb dermatan sulfates with chondroitinase AC-II. — To a solution of the polysaccharide (RC-20 or RC-30, 2 mg/0.2 mL of water) and enriched Tris buffer⁸, pH 8.0 (0.1 mL), was added chondroitinase AC-II (10 units/0.2 mL of water), and the mixture incubated for 3 h at 37°. After being heated in boiling water for 10 min, the mixture was centrifuged, and the supernatant solution was applied to a Sephadex G-200 column (1.5 × 90 cm) prepared in 0.15M sodium chloride. The column was eluted with the same solvent at a flow rate of 20 mL/h, and each fraction collected (2 mL) was analyzed for both uronic acid content and absorbance at 232 nm.

On a preparative scale, a solution of RC-20 or RC-30 dermatan sulfate (50 mg/3 mL of water) and enriched Tris buffer (pH 8.0, 1 mL) was incubated with chondroitinase AC-II (20 units/1 mL of water) for 16 h at 37°. After treating the incubation mixture as just described, the supernatant solution was chromatographed on a Sephadex G-200 column (2.5 × 86 cm) prepared in 0.15M sodium chloride. Fractions corresponding to the elution area to be isolated (for example, Peak 1 or 2 in Fig. 1b) were pooled, lyophilized, and desalted on a Sephadex G-25 column (2.5 × 80 cm) prepared in 10% ethanol and eluted with the same solvent. The solution was evaporated *in vacuo* at 35°, and lyophilized. The Peak 2 material of RC-20 or RC-30 was analyzed for 4,5-unsaturated disaccharide composition according to the procedure previously described¹. The Peak 1 material of RC-30 was digested with chondroitinase ABC according to the determination of uronic acid composition of the preceding paper¹ to obtain a value of A_{232} chondroitinase AC-II/ A_{232} chondroitinase ABC.

Fragmentation of rooster-comb dermatan sulfates with periodate oxidation, followed by alkali cleavage. — The polysaccharide fractions, RC-20 and RC-30, were oxidized and cleaved by the procedure described by Fransson *et al.*^{9,10}. A solution of the polysaccharide (100 mg) in 20mM sodium periodate and 50mM sodium citrate buffer (pH 3.0, 50 mL) was kept in the dark for 24 h at 4°. A 10% solution of mannitol (5 mL) was added, the mixture dialyzed against distilled water (5 L × 5) for 48 h, and the dialyzate lyophilized. The residue was dissolved in water (5 mL), and the pH adjusted to 12 with M sodium hydroxide. The solution was kept

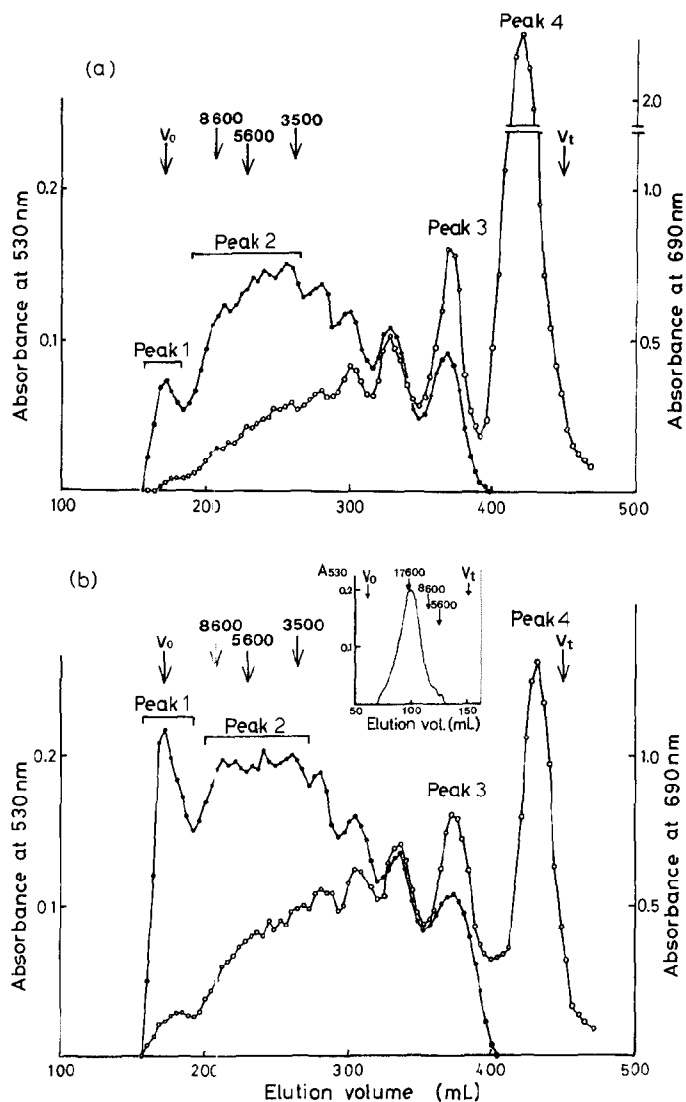


Fig. 2. Separation, on Sephadex G-50, of the products formed by periodate oxidation and alkali degradation of rooster-comb dermatan sulfates: (a) RC-20, and (b) RC-30. Each column was calibrated with Blue dextran (V_0), depolymerized chondroitin 6-sulfates (M_r 8600, 5600, and 3500), and 2M sodium chloride (V_t). The elution diagram, from Sephadex G-200, of Peak 1 material of RC-30 is shown in the inset of Fig. 2b. (—●—●—●—) $A_{530\text{nm}}$, and (—○—○—○—) $A_{690\text{nm}}$.

for 30 min at room temperature, made neutral with M hydrochloric acid, and applied to a Sephadex G-50 column (2.5×91 cm) prepared in 0.15M sodium chloride. The column was eluted with the same solvent at a flow rate of 20 mL/h. Each 4 mL-fraction was collected and analyzed for both uronic acid content and reducing power (Figs. 2a and b).

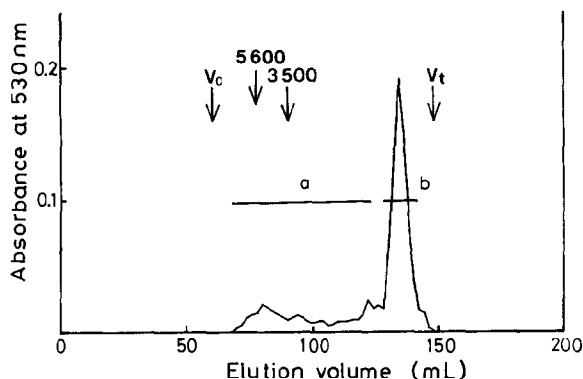


Fig. 3. Separation, on Sephadex G-50, of the digestion products, with chondroitinase AC-II, of the high-mol.-wt. fraction (Peak 1 of Fig. 2b) obtained by periodate oxidation of rooster-comb dermatan sulfate, RC-30. The column was calibrated with Blue dextran (V_0), depolymerized chondroitin 6-sulfates (M_r 5600 and 3500), and 2M sodium chloride (V_t). The horizontal bars indicate (a) higher oligo- and (b) di-saccharide areas.

Fractions constituting the peak near V_0 (Peak 1 in Fig. 2b; elution vol., 156–192 mL) were pooled, lyophilized, and desalted on a Sephadex G-25 column (2.5 \times 80 cm) by elution with 10% ethanol, and the desalted solution (pH 6.5) evaporated at 35° to remove ethanol and lyophilized. The residue was dissolved in 0.15M sodium chloride (1 mL; 1 mg/mL) and chromatographed on a Sephadex G-200 column (1.5 \times 83 cm), prepared in 0.15M sodium chloride and eluted with the same solvent. The column was calibrated with Blue dextran (V_0), depolymerized chondroitin 6-sulfates (M_r 17 600, 8600, and 5600), and 2M sodium chloride (V_t) (inset of Fig. 2b).

The ability of Peak 1 material to be degraded by chondroitinase AC-II was examined as follows. To a solution of this material (1 mg/0.1 mL of water) and enriched Tris buffer, pH 8.0 (0.05 mL), was added chondroitinase AC-II (5 units/0.1 mL of water), and the mixture incubated for 3 h at 37°. After being heated in boiling water for 10 min, the mixture was centrifuged, and the supernatant solution applied to a Sephadex G-50 column (1.5 \times 86 cm) prepared in 0.15M sodium chloride. The column was eluted with the same solvent at a flow rate of 20 mL/h, and each fraction collected (0.5 mL) was analyzed for uronic acid content (Fig. 3).

Fractions corresponding to a broad area at the rear of Peak 1 (Peak 2 in Fig. 2b; elution vol., 200–274 mL) were pooled, and Peak 2 material was isolated as described previously. The materials of Peaks 1 and 2 (each 100 μ g) were digested separately with chondroitinase ABC (0.2 unit in 20 μ L of water) in enriched Tris buffer, pH 8.0 (30 μ L), and with chondroitinase AC-II (0.5 unit in 20 μ L of water) in 50mM acetate buffer, pH 6.0 (30 μ L) for 3 h at 37°, respectively. After dilution of the reaction mixtures with 10mM hydrochloric acid to a final volume of 1.5 mL, the absorbance at 232 nm was measured, and the ratio between the values obtained from the two enzymic digests was calculated (Table I).

TABLE I

DEGRADATION WITH CHONDROITINASES AC-II AND ABC OF THE PRODUCTS OF PERIODATE OXIDATION OF RC-30^a

Degradation with chondroitinase	Peak ^b	
	1	2
AC-II	0.476	0.515
ABC	0.670	0.678
Ratio of AC-II to ABC value	0.71	0.76

^aMaterials from Peaks 1 and 2 (see legend to Fig. 2b). ^bA₂₃₂ for 100 µg of material oxidized.

RESULTS AND DISCUSSION

Fractions RC-20 and RC-30 were digested with chondroitinase AC-II according to the procedure of Saito *et al.*⁸ The gel-filtration diagrams, on Sephadex G-200 (Fig. 1a and 1b), of the digestion products showed a large peak (Peak 2; elution volume for RC-20, 145–160 mL; and for RC-30, 140–160 mL) of unsaturated disaccharide monosulfate, originating from the sequentially linked *N*-acetylchondrosine units, and a group of peaks differing in the degree of polymerization (elution vol. for RC-20, 60–145 mL; and for RC-30, 65–140 mL), which originate from the sequentially linked *N*-acetyldermosine units. The digestion products of RC-20 or RC-30 were fractionated on a preparative scale by Sephadex G-200 gel-filtration to isolate Peak-2 materials. Each of these was then subjected to analytical gel-filtration on Sephadex G-25 and paper chromatography, and was identified as Δ Di-4S (containing a negligible proportion of Δ Di-OS) (data not shown). The ratio of total peak area A₂₃₂, as measured by the elution diagrams Fig. 1a and 1b for RC-20 to that for RC-30, was found to be 0.56:1. Since the elution diagrams were obtained for the digestion products resulting from treatment of equal amounts of RC-20 and RC-30 with chondroitinase AC-II, the ratio (0.56:1) represents that of the contents of total *N*-acetylchondrosine units of RC-20 and RC-30. This result agrees well with the ratio¹ (0.55:1) of the D-glucuronic acid contents (22.6% for RC-20 and 41.1% for RC-30). Each of the A₂₃₂ area of Peaks 2 of RC-20 and RC-30 (representing the total amounts of the unsaturated disaccharide from the sequentially linked *N*-acetylchondrosine units) corresponds to 87% of the total A₂₃₂ peak areas, indicating a distribution of *N*-acetylchondrosine units very similar in RC-20 and RC-30; this ratio corresponds to the uronic acid composition. As indicated in the preceding paper¹, most of the *N*-acetylchondrosine units exist as clusters, as a cluster of 8 *N*-acetylchondrosine units between *N*-acetyldermosine units, corresponds to a content of unsaturated disaccharides and tetra- or higher oligosaccharides of 87.5 and 12.5%, respectively.

A small, distinct peak (Peak 1), near *K*_{av} 0.61 (elution vol., 114–126 mL), was observed for both Fractions RC-20 and RC-30 (Fig. 1a and 1b). In order to es-

timate the average M_r of Peak 1 material, Fraction RC-30 was digested with chondroitinase AC-II on a preparative scale, and Peak-1 material, isolated by gel-filtration of the digestion product, was digested with chondroitinase ABC. The ratio of the A_{232} value for the digestion product with chondroitinase AC-II to that with chondroitinase ABC was found to be 0.081–0.072:1. From this value, it was estimated that Peak-1 material contains, on average, 12–14 disaccharide units (11–13 *N*-acetyldermosine units plus 1 unsaturated disaccharide unit), assuming that the value of A_{232} due to the unsaturated uronic acid residue was not influenced by the length of the saccharide sequence linked to the residue*. From the average M_r of the RC-30 disaccharide unit (524), the M_r of Peak-1 material was calculated to be 6500–7300. This value agrees with the M_r range obtained by gel chromatography (Figs. 1a and 1b). Thus, Peak-1 material resulted from a polysaccharide chain having, on average, 11–13 sulfated *N*-acetyldermosine units between *N*-acetylchondrosine units. This structure seems characteristic for rooster-comb dermatan sulfate.

A large proportion of high-mol.-wt. products, in which no unsaturated uronic acid (A_{232}) was detected and which corresponds to 46% of the uronic acid area of the total digestion products, was eluted early (Fig. 1a; elution vol., 60–92 mL). Taking into consideration the sensitivity of the A_{232} measurement, it seems that most of the materials corresponding to this area derived from large *N*-acetyldermosine clusters containing nonreducing, terminal portions of RC-20. The M_r value estimated at $>17\ 600$ (>34 sulfated *N*-acetyldermosine units; Fig. 1a) suggests the presence of a few particularly large clusters having a size comparable to that of the original polysaccharide chain¹ of RC-20 ($M_r \sim 41\ 000$). Similarly, a material that may have derived from large *N*-acetyldermosine clusters was also observed for RC-30, but in far smaller proportions and derived probably from contaminating RC-20.

As expected, the proportions of tetra- or higher oligosaccharides (up to $M_r \sim 17\ 600$) carrying an unsaturated uronic acid residue (a relatively small proportion, as estimated from the A_{530} to A_{232} ratios) or not carrying the residue (relatively abundant) were small for RC-20 (40% of total uronic acid content) and large for RC-30 (59%) (see Fig. 1a, elution vol. 94–144 mL; and Fig. 1b, elution vol. 90–140 mL). These different proportions reflect the contents of D-glucuronic acid and are also related to the different chain structures at the nonreducing, terminal end.

Periodate oxidation of RC-20 and RC-30, followed by alkali degradation^{9,10}, gave fragments that were chromatographed on Sephadex G-50 (Figs. 2a, 2b). A substantial proportion of the fragments from RC-20 was included in the gel, and the smallest product arising from this degradation of the sequentially linked *N*-acetyldermosine monosulfate units [$\text{GalNAc-O-C(CHO)=CH-CO}_2\text{H}$] gave a large peak (Peak 4, Fig. 2a; elution vol. 400–460 mL). Peak 3 (Fig. 2a; elution vol. 350–390 mL) contained $(\text{GalNAc} \rightarrow \text{UA}) \rightarrow \text{GalNAc-O-C(CHO)=CH-CO}_2\text{H}$ (UA:

*The method for measurement of the GlcA-to-IdoA ratio of dermatan sulfate using digestion with chondroitinases AC-II and ABC is based on this assumption.

GlcA or IdoA sulfate), which originated from D-glucosyluronic acid→GalNAc 4-sulfate or L-idosyluronic acid sulfate→GalNAc 4-sulfate units between monosulfated *N*-acetyldermosine units. The total amount of uronic acid in the peak areas of Fig. 2a corresponds to 33% of the uronic acid present in the starting fraction RC-20. This value is approximately equal to the sum¹ of D-glucuronic acid (23%) and Δ Di-diS (7%)¹. A similar agreement between these corresponding values was observed for RC-30. Thus the disaccharide giving Δ Di-diS contains L-iduronic acid sulfate.

In RC-30, the formation of GalNAc-O-C(CHO)=CH-CO₂H (Peak 4, Fig. 2b) was less significant than in RC-20, but the proportion of high-molecular-weight products (Peaks 1 and 2; elution vol., 156–272 mL) was large, and the total uronic acid content of the Peak-1–3 area corresponded to 56% of the uronic acid present in starting RC-30 (in contrast to 33% for RC-20). In particular, Peak 1 of RC-30 (Fig. 2b), which contains the large-size fragments excluded from the Sephadex G-50 gel, was much greater than Peak 1 of RC-20 (Fig. 2a). The different elution diagrams for RC-30 and RC-20 results from the higher content in D-glucuronic acid (41%) and higher content in disaccharide disulfate (13%) in RC-30 than in RC-20 (23 and 7%, respectively), which suggests that RC-30 is rich in large-size clusters of sulfated (or nonsulfated) *N*-acetylchondrosine and *N*-acetyldermosine disulfate units.

The structure of the large-size clusters of RC-30 was elucidated by digesting, with chondroitinases AC-II and ABC, the fractions of higher M_r (Fig. 2b, Peak 1 and following broad Peak-2-fraction of $M_r > 3500$) and by examining, on Sephadex G-50, the product of digestion of Peak 1 material with chondroitinase AC-II (see Table I and Fig. 3). The polysaccharide chain of these higher-molecular-weight products is constituted preponderantly of *N*-acetylchondrosine monosulfate units with several *N*-acetyldermosine disulfate units distributed within the chain.

For Peak 4 material of RC-20, the proportion of uronic acid, oxidized to give GalNAc-O-C(CHO)=CH-CO₂H, relative to the total uronic acid of the starting material is 67% (100 – 33%), which is approximately equal to the proportion of nonsulfated IdoA, 70% [total IdoA (77) – Δ Di-diS (7)]. For RC-30, these values are 44 (100 – 56) and 45% (59 – 14), nearly equal. Thus, it is highly likely that most *N*-acetyldermosine disulfate units that give Δ Di-diS are distributed within the *N*-acetylchondrosine sulfate clusters rather than within the *N*-acetyldermosine monosulfate clusters.

In summary, in agreement with our previous results¹, both RC-20 and RC-30 fractions have many *N*-acetylchondrosine or *N*-acetyldermosine clusters of various sizes, but they differ in the proportions of the uronic acids and in the content of disaccharide disulfate units. In addition, the present work, has shown: (a) RC-20, but not RC-30, contains many *N*-acetyldermosine sulfate clusters of a large size ($M_r \sim 17\,600$ or more) at the nonreducing, terminal end. (b) Both fractions have an *N*-acetyldermosine sulfate cluster of M_r 6500–7300 within the chain. This seems characteristic of rooster-comb dermatan sulfate. (c) RC-30 contains more D-

glucuronic acid than RC-20. In both fractions, most *N*-acetylchondrosine sulfate units form clusters. Several of these have a large size (M_r ~17 600, ~34 disaccharide sulfate units) in RC-30, but not in RC-20. In both fractions, many of the *N*-acetylchondrosine sulfate clusters have a M_r of ~3500–9000. (d) In RC-30, several *N*-acetyldermosine disulfate units (13%) may be linked to *N*-acetylchondrosine sulfate units or clusters, suggesting that, in both fractions, most *N*-acetylchondrosine sulfate clusters having a large molecular size exist as a copolymer with *N*-acetyldermosine disulfate units.

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